Mid-Atlantic Environmental Consultants

SALEM UNITED METHODIST CHURCH
CLASSROOM # 5

MOLD REPORT

MID ATLANTIC JOB NUMBER:
MCOM-23-33

OCTOBER 2023

PREPARED FOR:

SALEM UNITED METHODIST CHURCH
350 MANOR ROAD
WEXFORD, PA 15090

PREPARED BY:

MID ATLANTIC ENVIRONMENTAL CONSULTANTS, INC.
5320 N. PIONEER ROAD
GIBSONIA, PA 15044
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Mid-Atlantic Environmental Consultants

5320 N. Pioneer Road
Gibsonia, PA 15044
Phone: 724-444-3460
Fax: 724-444-3463
Email: midatlantic@zoominternet.net

October 23, 2023

Salem United Methodist Church
350 Manor Road
Wexford, PA 15090

Attn: (Reverend) Mrs. Stephanie Gottschalk

Re: Mold Testing

To Whom It May Concern:

On Tuesday, October 17th, 2023, Mid Atlantic Environmental Consultants, Inc. performed mold air quality testing as requested by Reverend Stephanie Gottschalk, at the Salem United Methodist Church located at 350 Manor Road in Wexford, Pennsylvania. The purpose of this visit was to perform mold air sampling to determine the fungal ecology inside classroom #5.

We appreciate the opportunity to assist you with this project. Should you have any further questions or concerns, please do not hesitate to contact Mid Atlantic by phone at: (724) 444-3460; or by e-mail at: midatlantic@zoominternet.net.

Sincerely,

Joseph D. Pillart
Certified Indoor Environmentalist

SALEM UNITED METHODIST CHURCH – CLASSROOM #5

Mid Atlantic Environmental Consultants, Inc. was retained to conduct mold air sampling at the Salem United Methodist Church located at 350 Manor Road in Wexford, Pennsylvania. Mid Atlantic representative Mr. Joseph Pillart performed mold sampling activities. Mr. Pillart is a Certified Indoor Environmentalist (CIE) certified by the American Council for Accredited Certifications (ACAC).

Mold air sampling was conducted inside classroom #5 to check the fungal ecology inside the classroom after a water leak into the classroom from a roof leak. The roof leak was already repaired, and the clean-up of the classroom was done before the site visit. A limited visual inspection of the sampled area found suspect black-like mold on the ceiling above the drop ceiling tiles. A moisture meter was used to check the carpet for moisture, and it was found to be dry during the site visit.

A total of two (2) mold air spore trap samples were collected, including one from the outside to use as a reference sample. One (1) mold surface swab was also collected. SanAir Technologies Laboratory in N. Chesterfield, Virginia performed the analysis of all samples collected. SanAir analysis procedures are performed in an AIHA EMPAT accredited laboratory and by the most current parameters set forth by the American Conference of Governmental Industrial Hygienists Bioaerosol Guidelines. Refer to Appendix A for further information.

MOLD AIR SPORE TRAP SAMPLE RESULTS FOR: OCTOBER 17TH, 2023
(Refer to Appendix A)

The results are reported as total meaning they include both viable and non-viable fungal spores.

M-1 - was collected from Classroom #5; revealing a total fungal spore count of 973; the most predominant fungi recovered was *Basidiospores.*

M-2 - was collected Outside to serve as a reference; revealing a total fungal spore count of 56,040; the most predominant fungi recovered was *Basidiospores.*

The inside air sample M-1 (Classroom #5) has a very low fungal counts and are comparable to the outside reference sample, indicating acceptable fungal ecology in the classroom area according to EPA and other industry guidelines.

Note: Smuts and Myxomycetes are molds that are commonly identified outside. They commonly grow on logs, grasses, weeds, and plants. Myxomycetes can grow inside but require a great deal of moisture to develop. Smuts are not usually found indoors. If found indoors it's typically from outside air infiltration. These are not considered hazardous molds. These two kinds of molds are put under the exact category since the spores look similar and cannot be distinguished independently under a microscope. The presence of Basidiospores usually indicates outside air infiltration. Indoor building materials usually do not support their growth.

All sample results represent data that was collected at an exact time - during certain indoor air quality conditions.

Mold Surface Swab Sampling:

Mold surface swab sampling was conducted by utilizing a Health Link Transporter sterile swab. Approximately a one square inch surface was sampled. The results are reported as total, meaning they include both viable and non-viable fungal spores.

Sample S-1 was collected from Classroom #5 - Ceiling revealing: **NO FUNGI DETECTED.**

The mold surface swab sample results confirms NO MOLD GROWTH on the sampled area. No further action is warranted.

CONCLUSION

Most guidelines such as those of American Industrial Hygienists Association (AIHA) American Conference of Governmental Industrial Hygienists (ACGIH), the United States Environmental Protection Agency (USEPA) and the Institute of Inspection Cleaning and Restoration Certification (IICRC S520) do not recommend strict numerical values as indicators of contamination. Instead, the accepted standard for the industry is that the inside air should be as good (equal) or better than outside air, both in total spore numbers and in representative genera/species. The only caution to this is when environmental conditions, such as rain or those encountered in the winter; prevent accurate representation on indigenous microbial populations. There are no specific regulations governing surface microbiological contamination of airborne microbiological contaminants in indoor air (bioaerosols). This is in part due to the many variables involved with sampling for microorganisms, dramatic fluctuations in background levels of microorganisms, lack of agreement between researchers about what constitutes a "problem situation" and an overall lack of industry experience in interpreting microbiological laboratory data.

None of the information contained in this report should be construed as medical advice or a call to action for evacuation. Any decision relative to medical significance should be made by a qualified physician.

Refer to the appendices for further information.

Appendix A—Sampling Information / Laboratory Analysis and Report
—Mold Sampling Data—October 17th, 2023

Appendix B—Project Photos

Appendix C—Accreditation

Should you have any further questions, feel free to contact our office at (724) 444-3460.

Appendix A – Sampling Information / Laboratory Analysis and Report Mold Sampling Data

SanAir Technologies Laboratory

The Identification Specialists

Analysis Report
prepared for
Mid Atlantic Environmental Consultants Inc.

Report Date: 10/20/2023

Project Name: Room - 5

Project #: Salem UMC Pre-School

SanAir ID#: 23057995

10501 Trade Court | North Chesterfield, Virginia 23236
888.895.1177 | 804.897.1177 | fax: 804.897.0070 | IAQ@SanAir.com | SanAir.com

SanAir Technologies Laboratory

SanAir ID Number
23057995
FINAL REPORT
10/20/2023 10:09:10 AM

Name: Mid Atlantic Environmental Consultants Inc.
Address: 5320 North Pioneer Road
Gibsonia, PA 15044
Phone: 724-444-3460

Project Number: Salem UMC Pre-School
P.O. Number: MCOM-23-33
Project Name: Room - 5
Collected Date: 10/17/2023
Received Date: 10/18/2023 10:40:00 AM

Dear Joe Pillart,

We at SanAir would like to thank you for the work you recently submitted. The 3 sample(s) were received on Wednesday, October 18, 2023 via UPS. The final report(s) is enclosed for the following sample(s): M-2, M-1, S-1.

These results only pertain to this job and should not be used in the interpretation of any other job. This report is only complete in its entirety. Refer to the listing below of the pages included in a complete final report.

Sincerely,

L. Claire Macdonald

L. Claire Macdonald
Microbiology Laboratory Manager
SanAir Technologies Laboratory

Final Report Includes:
- Cover Letter
- Air Cassette Analysis
- Direct ID Analysis
- Disclaimers and Additional Information

Sample conditions:
- 3 samples in Good condition.

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FINAL REPORT
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Analyst: Willis, Madeline

Air Cassette Analysis

ND = None Detected. Blank spaces indicate no spores detected.

SanAir ID Number	23057995-001			23057995-002		
Analysis Using STL	105C			105C		
Sample Number	M-1			M-2		
Sample Identification	Class Room #5			Outside Reference		
Sample Type	Air Cassette - Air-O-Cell			Air Cassette - Air-O-Cell		
Volume	75 Liters			75 Liters		
Analytical Sensitivity	13 Count/M³			13 Count/M³		
Background Density	2+			1+		
Other	**Raw Count**	**Count/M³**	**%**	**Raw Count**	**Count/M³**	**%**
Dander	495	6600	n/a	20	267	n/a
Fibers	56	747	n/a	3	40	n/a
Mycelial Fragments	3	40	n/a	2	27	n/a
Pollen				1	13	n/a
Fungal Identification	**Raw Count**	**Count/M³**	**%**	**Raw Count**	**Count/M³**	**%**
Ascospores	5	67	7	58	773	1
Aspergillus/Penicillium	5	67	7	3	40	< 1
Basidiospores	44	587	60	4090	54533	97
Cladosporium species	5	67	7	49	653	1
Curvularia species	2	27	3			
Polythrincium species				1	13	< 1
Rusts	1	13	1			
Smuts/Myxomycetes	11	147	15	2	27	< 1
TOTAL	**73**	**973**		**4203**	**56040**	

Signature: Madeline Willis Date: 10/20/2023 Reviewed: Jonathan Wilson Date: 10/20/2023

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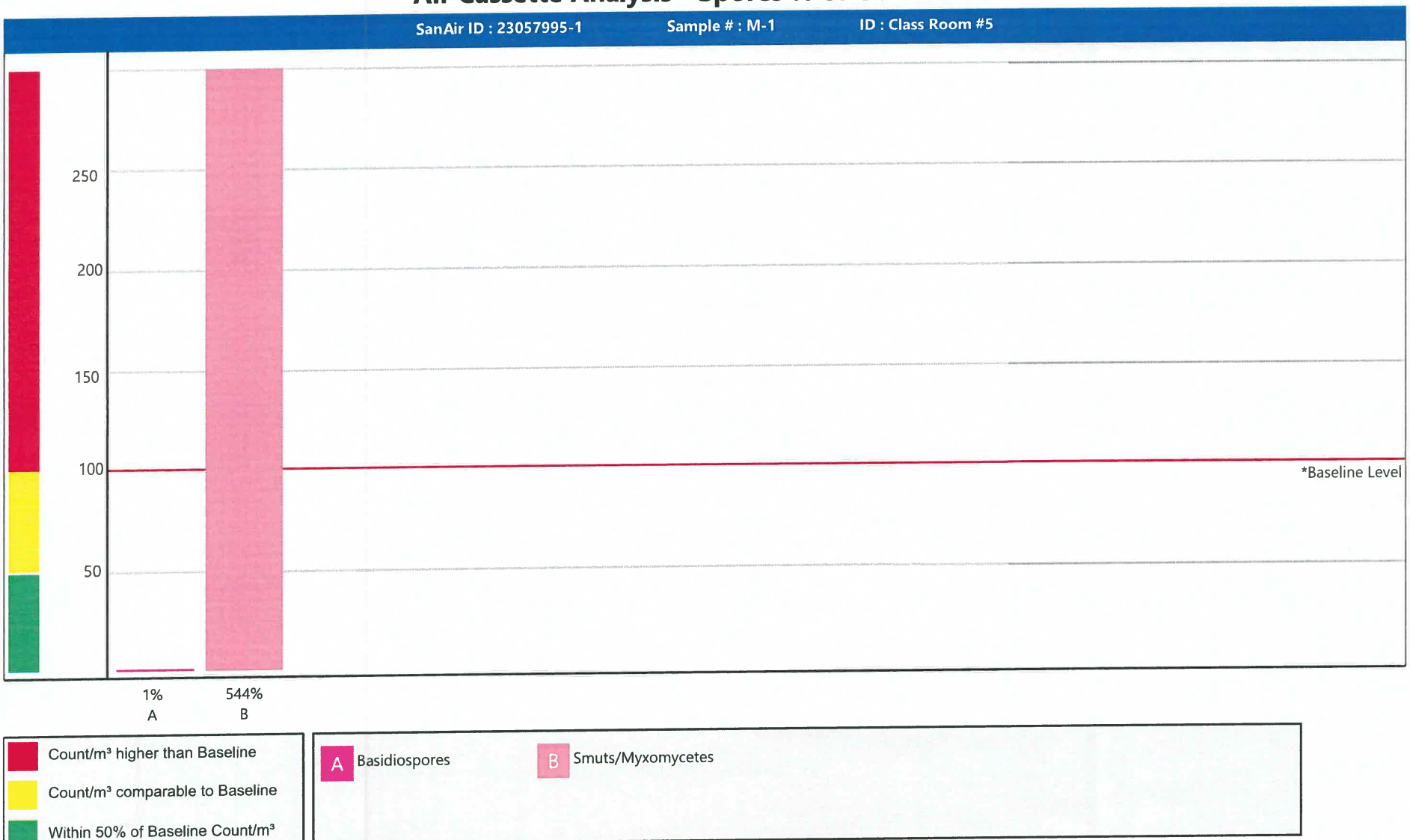
Analyst: Willis, Madeline

Air Cassette Analysis - Spores % of Outside Air



*The Baseline Level (100%) represents the average baseline sample counts. Counts above the baseline may indicate higher than expected levels of a given result.

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Analyst: Zhang, Ph.D, Richard

Direct Identification Analysis

SanAir ID: 23057995-003 Sample #:S-1 Class Room #5 - Ceiling

D1 - Direct Identification Analysis on Surface Swab using STL 104

Direct ID of Mold

Fungi	Estimated Amount
No Fungi Detected	

Estimated Amount	Indication of Growth	Evidence of Mycelial Fragments/Conidiophores
Rare	Not Likely	None
Light	Possible	Some, 10 to 25% of Tape Covered
Moderate	Probable	Abundant, 25 to 50% of Tape Covered
Heavy	Significant	Throughout, 50 to 100% of Tape Covered

*Refer to additional information page for further details

Signature: Date: 10/20/2023

Reviewed: Date: 10/20/2023

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Organism Descriptions

The descriptions of the organisms presented are derived from various reference materials. The laboratory report is based on the data derived from the samples submitted and no interpretation of the data, as to potential, or actual, health effects resulting from exposure to the numbers of organisms found, can be made by laboratory personnel. Any interpretation of the potential health effects of the presence of this organism must be made by qualified professional personnel with first hand knowledge of the sample site, and the problems associated with that site.

Dander - Comprised of human and/or animal skin cells. Counts may be higher in carpeted rooms and in rooms with more traffic.
Health Effects: May cause allergies.

Fibers - This category can include clothing, carpet, and insulation fibers.

Mycelial Fragments - A mycelium (plural = mycelia) is the "body" of a fungus. It is a collective term for hyphae (singular = hypha), which are the tubular units of the mycelium usually composed of chitin. The terms hyphae and mycelial fragments are used interchangeably. [This information was referenced from the mycology text "The Fifth Kingdom"]In some cases a fungal identification cannot be obtained due to lack of sporulation. Only the mycelial fragments are present, and cannot be identified without the distinguishing characteristics of the spores or the structures they grow from.
Health Effects: Allergic reactions may occur in the presence of spores (conidia) or mycelial/hyphal fragments.

Pollen - Produced by trees, flowers, weeds and grasses. The level of pollen production can depend on water availability, precipitation, temperature, and light. Pollen is usually dispersed by either insects or the wind.
Health Effects: Mostly effects the respiratory tract with hay fever symptoms but has also been shown to trigger asthma in some people.

Ascospores - From the fungal Subphylum Ascomycotina. Ascospores are ubiquitous in nature and are commonly found in the outdoor environment. This class contains the "sac fungi" and yeasts. Some ascospores can be identified by spore morphology, however; some care should be excercised with regard to specific identification. They are identified on tape lifts and non-viable analysis by the fact that they have no attachment scars and are sometimes enclosed in sheaths with or without sacs. Ascomycetes may develop both sexual and asexual stages. Rain and high humidity may help asci to release, and dispurse ascospores, which is why during these weather conditions there is a great increase in counts.
Health Effects: This group contains possible allergens.

Aspergillus/Penicillium - These spores are easily aerosolized. Only through the visualization of reproductive structures can the genera be distinguished. Also included in this group may be spores of the genera Acremonium, Phialophora, Verticillium, Paecilomyces, Talaromyces etc. Small, round to ovoid spores of this group lack the necessary distinguishing characteristics when seen on non-viable examination.
Health Effects: Can cause a variety of symptoms including allergic reactions. Most symptoms occur if the individual is immunocompromised in some way (HIV, cancer, etc).

Basidiospores - From the Subphylum Basidiomycotina which contains the mushrooms, shelf fungi, and a variety of other macrofungi. They are saprophytes, ectomycorrhizal fungi or agents of wood rot, which may destroy the structure wood of buildings. It is extremely difficult to identify a specific genera of mushrooms by using standard culture plate techniques. Some basidiomycete spores can be identified by spore morphology; however, some care should be exercised with regard to specific identification. The release of basidiospores is dependant upon moisture, and they are dispersed by wind.
Health Effects: Many have the potential to produce a variety of toxins. Members of this group may trigger Type I and III fungal hypersensitivity reactions. Rarely reported as opportunistic pathogens.

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Cladosporium species - The most commonly identified outdoor fungus. The outdoor numbers are reduced in the winter and are often high in the summer. Often found indoors in numbers less than outdoor numbers. It is commonly found on the surface of fiberglass duct liner in the interior of supply ducts. A wide variety of plants are food sources for this fungus. It is found on dead plants, woody plants, food, straw, soil, paint and textiles. Often found in dirty refrigerators and especially in reservoirs where condensation is collected, on moist window frames it can easily be seen covering the whole painted area with a velvety olive green layer.
Health Effects: It is a common allergen. It can cause mycosis. Common cause of extrinsic asthma. Acute symptoms include edema and bronchiospasms, chronic cases may develop pulmonary emphysema. Illnesses caused by this genus can include phaeohyphomycosis, chromoblastomycosis, hay fever and common allergies.
References: Flannigan, Brian, Robert A. Samson, and J. David Miller, eds. Microorganisms in Home and Indoor Work Environments. London and NY: Taylor & Francis, 2001.de Hoog, G.S. et al. Atlas of Clinical Fungi. 4th ed. Foundation Atlas of Clinical Fungi. 2020

Curvularia species - Curvularia is found on plant material and is considered a saprobe. It has also been isolated from dust samples and from wallpaper.
Health Effects: It has been reported to be a cause of allergic fungal sinusitis. It may cause corneal infections, mycetoma and infections in immune compromised hosts.
References: de Hoog, G.S. et al. Atlas of Clinical Fungi. 4th ed. Foundation Atlas of Clinical Fungi. 2020

Polythrincium species - This fungus is often associated with leaves and other plant material. There are no reports of any clinical significance or allergenic properties.
References: Ellis, Martin B., Ellis, Pamela, Microfungi on Land Plants: An Identification Handbook. England, The Richmond Publishing Co. Ltd., 1997.

Rusts - From the group Uredinales, called Rusts due to the color of the spores, which are known for causing disease in plants.

Smuts/Myxomycetes - Smuts and Myxomycetes are parasitic plant pathogens. They are typically grouped together due to their association with plants, the outdoors and because they share similar microscopic morphology.
Health Effects: Can produce type I fungal hypersensitivity reactions.
References: Martin, G.W., C.J. Alexopoulos, and M.L. Farr. The Genera of Myxomycetes. Iowa City, Iowa: University of Iowa Press, 1983.

Additional Information

Air Cassette Analyses

Air cassette reports indicate the genus and concentration of viable (living) and non-viable mold spores detected on the slide (A2 Analysis). Whether or not these spores are viable cannot be determined using this type of analysis. However, keep in mind that spores can remain allergenic even after cellular death. Other possible allergens include dander, pollen and fibers which are included in air cassette reports for the A1 Analysis. A1 and A2 analyses are performed on several types of air cassettes. Light microscopy at a 400 to 1000x magnification is used for air cassette sample analysis. SanAir always analyzes 100% of the impacted slide.

Explanation of Background Densities

The background density of an air cassette aids in the overall interpretation of results as it indicates the level of background debris present (e.g. dander, pollen, fibers, insect parts, soot, fly ash, etc.). Excessive background debris may mask the presence of fungal spores thereby reducing the accuracy of the count. It may also serve as an alert that the volume of air pulled was too high or too low. The following table explains background densities.

Air Cassette Density	Amount of Particulate on Slide	Explanation
1	Insignificant	Should not skew any counts
1+	Low	Should not skew any counts
2	Low to Moderate	Should not skew any counts
2+	Moderate to High	May cause occlusion of small spores
3	High	May cause occlusion of small to medium spores
3+	Very High	Will cause occlusion of spores
4	Overloaded	Level of particulate too high to perform analysis

A Note About the Fungal Spores

In some instances certain groups of fungi cannot be identified due to a lack of distinguishing characteristics. These fungi will be categorized as "non-specified spores" on the final report.

The genera *Aspergillus* and *Penicillium* are typically composed of small, round spores that are difficult to distinguish from each other; therefore, they are grouped into the category *Aspergillus / Penicillium*. Other fungi that produce spores of similar characteristics may also be placed into this category, including *Paecilomyces*, *Talaromyces*, and *Trichoderma*, among others.

Stachybotrys and *Memnoniella* spores are coated with a sticky "slime" layer that may inhibit aerosolization.

Any genus of fungi detected on an air cassette with a high raw count (i.e. exceeding 500 spores) may be estimated. Any estimate higher than 12,000 spores will be reported as >12,000.

Understanding the Air Cassette Report

Each sample has 3 columns of information provided. The left is the raw count which is the number of spores for that fungal type detected on the trace. The middle column is the count per cubic meter (Count/m^3) which is the raw count converted based on the total volume pulled for that sample. It represents the number of spores that should be expected in a cubic
meter of air from the location in question *if* the spores were distributed evenly throughout the air. This column is helpful for interpreting results when the samples were pulled at different total volumes. In other words, the raw count of a cassette pulled at 75 liters should not be compared to the raw count of a cassette pulled at 150 liters because there may be higher counts associated with the higher volume. By comparing the "Count/m^3" columns the difference in volumes are accounted for.

The analytical sensitivity is the lowest spore count detectable with reasonable certainty, and it is calculated this way using a raw count of one. Keep in mind there are 1,000 liters in a cubic meter.

1 x (1,000 / Total Volume in Liters)

How to calculate the count per cubic meter:

Raw Count x (1,000 / Total Volume in Liters)

The last column on the right shows the percentage for which each spore type comprised the total spore count.

Understanding the Air Cassette Graph (If included in the final report)

The graph is a visual representation of the baseline sample (usually the outdoor air sample) compared individually against each indoor sample. Each spore type found on the indoor sample is compared to what was found outdoors per cubic meter.

The graph shows the percentile representation of each indoor spore count derived by dividing the indoor Count/m^3 by the outdoor Count/m^3. If the percentage is below 50% of the outside count, then the bar is below 50 on the chart, which corresponds to "Within 50% of Baseline Count/m^3." If the percentage is between 50 and 100%, then the bar on the chart will stop between 50 and 100, which corresponds to "Count/m^3 comparable to Baseline." If the percentage is greater than 100%, then the bar will be above 100 on the chart, which corresponds to "Count/m^3 higher than Baseline."

Each organism is given a threshold level for the Count/m^3. If this threshold level is not met in an inside sample, then the organism will not be graphed on the chart. This is used to prevent the graph from showing every spore type that is commonly found outside and doesn't typically indicate a possible moisture problem inside. For example, most common outdoor spores (e.g. ascospores, basidiospores, and *Cladosporium*) have a threshold level of 100. Therefore, in order to show up on the chart, the inside Count/m^3 must be above 100. On the other hand, fungi that may indicate water damage (e.g. *Stachybotrys*, *Ulocladium*, *Chaetomium*, *Memnoniella*, etc.) are given lower threshold levels. These fungi have a higher water activity value and therefore require more moisture to grow. *Stachybotrys* and *Chaetomium* have threshold values of 14 and 30, respectively, as even a low count of those types of spores may indicate an issue with excess moisture.

Keep in mind that this graph is to be used only as a tool in the inspection of a building. Visual examination and knowledge of water damage, past remediation, and weather conditions, among other elements, is essential in the decision regarding the indoor air quality of a building.

Assistance with Remediation Projects

more information pertaining to interpretation of results is available on our website www.sanair.com

For assistance in a remediation project you may consult the Institute of Inspection, Cleaning and Restoration Certification's (IICRC) S500 and S520 protocols. The S500 is a reference guide for water-damage restoration and the S520 pertains specifically to mold remediation. Other standards and guidelines regarding Indoor Air Quality that may assist in remediation projects:

AIHA (Recognition, Evaluation, and Control of Indoor Mold)
AIHA (The Facts About Mold)
NADCA (ACR 2006)
IESO (Standards of Practice for the Assessment of Indoor Air Quality)
EPA (Mold Remediation in Schools and Commercial Buildings)
New York City Department of Health and Mental Hygiene (Guidelines on Assessment and Remediation of Fungi in Indoor Environments)

Disclaimer

SanAir Technologies Laboratory does not make contamination corrections to reports based upon analysis of laboratory and/or field blanks.

This report is the sole property of the client named on the SanAir Technologies Laboratory chain-of-custody. Results in the report are confidential information intended only for the use by the customer listed on the chain of custody (COC). Neither results nor reports will be discussed with or released to any third party without our clients' written permission. Final reports cannot be reproduced, except in full, without written authorization from SanAir. This report and any information contained within shall not be edited, altered, or modified in any way by any persons or agencies receiving, viewing, distributing, or otherwise possessing a copy of this final report. The laboratory reserves the right to perform amendments to any finalized report, of which shall supersede and make obsolete any previous editions. Such changes, modifications, additions, or deletions shall be effective immediately upon notice thereof, which may be given by means including but not limited to posting on the SanAir client portal website, electronic or conventional mail, or by any other means. The information provided in this report applies only to the samples submitted and is relevant only for the date, time, and location of sampling. The accuracy of the results of the analysis is dependent upon the method of sample procurement and information provided by the client on the COC. SanAir assumes no responsibility for the method of sample procurement. SanAir assumes no responsibility for information provided by the client on the COC such as project number, project name, collection dates, po number, special instructions, samples collected by technician name, sample numbers, sample identifications, sample type, selected analysis type, flow rate, total volume or area, and start stop times that may affect the validity of the results in this report. Evaluation reports are based solely on the sample(s) in the condition in which they arrived at the laboratory and on the information provided by the client on the COC. Sample(s) were received in good condition unless otherwise noted on the report. SanAir assumes no responsibility or liability for the manner in which the results are used or interpreted. SanAir will not provide any opinion on the safety of a building as visual inspection and knowledge of water damage, past remediation and weather conditions during sampling, among other elements, is essential in this decision. All samples are disposed of after 90 days unless otherwise requested by the client. SanAir is accredited by AIHA LAP, LLC in the EMLAP program. Refer to our accreditation certificate and scope or www.aihaaccreditedlabs.org for an up to date list of the Fields of Testing for which we are accredited.

This report does not constitute nor shall be used by the client to claim product, process, system, or person certification, approval, or endorsement by AIHA LAP, LLC, NVLAP, NELAC, NIST and/or any other U.S. governmental agencies; and may not be accredited by every local, state and federal regulatory agency.

LELAP Lab ID#05088

AIHA LAP, LLC Lab ID: LAP-162952

Additional Information

Direct Identification Analyses

Direct identification analyses can be performed on tape, bulk, dust and swab samples. Direct identification reports indicate the evidence of possible active growth for each genus of fungi present. Whether or not these spores are viable or nonviable cannot be determined using this type of analysis; the sample would have to be cultured in order to determine viability. Keep in mind that this report can only be inferred for the exact spot in which the sample was taken. Light microscopy at a 400 to 1000x magnification is used for direct identification analysis.

It is encouraged to include a blank tape sample in order to check for contamination during sampling or shipment. Be sure to check the expiration date of any tape. It is recommended not to use expired tapes as the gel on the slide deteriorates thereby losing the tackiness necessary to retain fungi.

The genera *Aspergillus* and *Penicillium* are typically composed of small, round spores that are difficult to distinguish from each other without the presence of intact conidiophores (structures from which spores are formed and released). In this case, they are grouped into the category *Aspergillus / Penicillium*. Other fungi that produce spores of similar characteristics to *Aspergillus* and *Penicillium* may also be placed into this combined category in the absence of intact conidiophores (e.g. *Paecilomyces*, *Gliocladium*, *Trichoderma*, etc.).

NOTE: Swabs are not the best media to use for direct analyses as all organisms may not be recovered intact, if at all, when analyzed.

NOTE: Tapes should not be overloaded with debris as that may occlude fungi.

D1 Analysis: Fungal Identification with "Evidence of Growth" Description

Results for the direct identification analysis describe the amount of evidence indicating possible fungal growth. The presence of associated mycelial fragments and conidiophores help the analyst to determine which description to use: rare, light, moderate, or heavy. Please refer to the following table for interpretation of direct identification results.

Estimated Amount	Indication of Growth	Evidence of Mycelial Fragments / Conidiophores
Rare	Not Likely	None
Light	Possible	Some, 10 to 25% of Tape Covered
Moderate	Probable	Abundant, 25 to 50% of Tape Covered
Heavy	Significant	Throughout, 50 to 100% of Tape Covered

Disclaimer

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SanAir Technologies Laboratory, Inc.

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Microbiology Chain of Custody

SanAir ID Number: 23057995

Mid Atlantic Environmental Consultants Inc.	Project ~~Number~~: Salem United Methodist Church Pre-School	Phone #:
5320 North Pioneer Road	Project Name: Room - 5	Phone #: 724-444-3460
Gibsonia, PA 15044	Date Collected: 10-17-23	Fax #:
Samples Collected By: Joe Pillart	P.O. Number: MCOM-23-33	Email: midatlanticq zoominternet.net

Sample Types		Analysis Types	Turn Around Time
AC	Air Cassette	A1 - Identification and Enumeration of Fungal spores, plus total dander, fiber, and pollen count	Hours 3/6/24/48-Std
		A2 - Identification and Enumeration of Fungal spores only	Hours 3/6/24/48-Std
T B S*	Tape Bulk Swab*	D1 - Direct Identification of Fungi	Hours 3/6/24/48-Std
		D2 - Direct Identification of Mites, Insects, Pollen, etc.	Hours 3/6/24/48-Std
AP B S	Air Plate Bulk Swab	C1 - Culture Identification and Enumeration of Fungi only	5-10 Days
		C2 - Culture Identification and Enumeration of Bacteria only	2-4 Days
		C3 - Culture Identification and Enumeration of Fungi and Bacteria	5-10 Days
		C4 - Culture Identification and Enumeration of Thermophilic Bacteria with C2 or C3 analysis	2-4 or 5-10 Days
W	Water	L1 – Culture Identification and Enumeration of *Legionella sp.*	7-10 Days
D	Dust	M1 – Dust Mite Allergen Test	Hours 3/6/24/48-Std

SanAir Technologies Laboratory offers speciation by PCR. Please call for details and pricing.

Sample #	Sample Identification	Sample Type	Analysis Type(s)	Turn Around Time	Total Volume (L) or Area (in^2)	Time Start – Stop
M-1	Class Room # 5	AC	A-1	STD	75L	N/A
M-2	Outside Reference	AC	A-1	STD	75L	N/A
S-1	Class Room # 5 - Ceiling	S*	D-1	STD	1sq inch	N/A

Special Instructions	

Relinquished by	Date	Time	Received by	Date	Time
[signature]	10-17-23	1300 UPS -	[signature]	10-18-23	10:40 Am

Unless scheduled, the turn around time for all samples received after 3 pm Friday will begin at 8 am Monday morning. Weekend or Holiday work must be scheduled ahead of time and is charged 150% of analytical rate.

*Although we allow Direct Identification from a swab sample, best results are received from tape samples.

Appendix B - Photographic Documentation

Mid-Atlantic Environmental Consultants

PHOTOGRAPHIC RECORD

SALEM UNITED METHODIST CHURCH - CLASSROOM #5

Photo #1:	Salem United Methodist Church

Photo #2:	Calibration of Pump – 15 lpm

Mid-Atlantic Environmental Consultants

PHOTOGRAPHIC RECORD

SALEM UNITED METHODIST CHURCH - CLASSROOM #5

Photo #3:	Mold Air Sample M-1 – Classroom #5

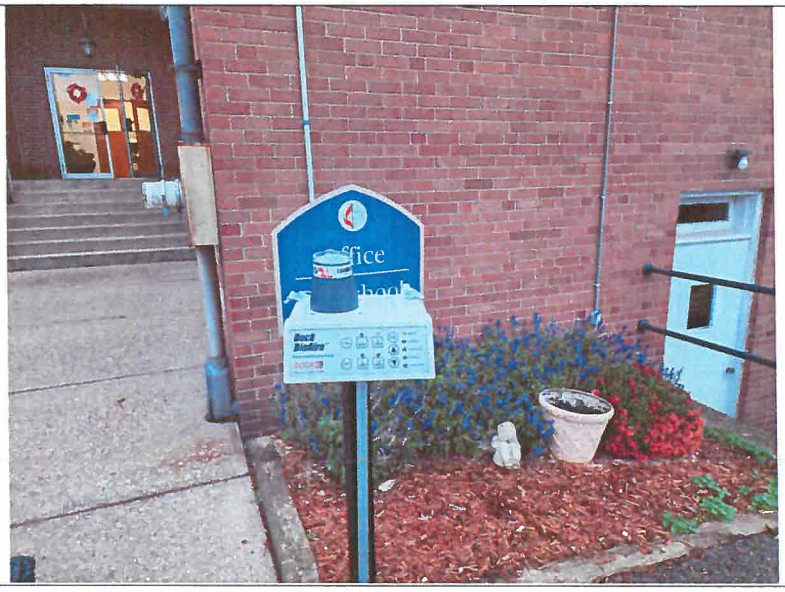

Photo #4:	Mold Air Sample M-2 – Outside Reference Sample

Mid-Atlantic Environmental Consultants

PHOTOGRAPHIC RECORD

SALEM UNITED METHODIST CHURCH - CLASSROOM #5

Photo #5:	Mold Surface Sample S-1 – Classroom #5 Ceiling

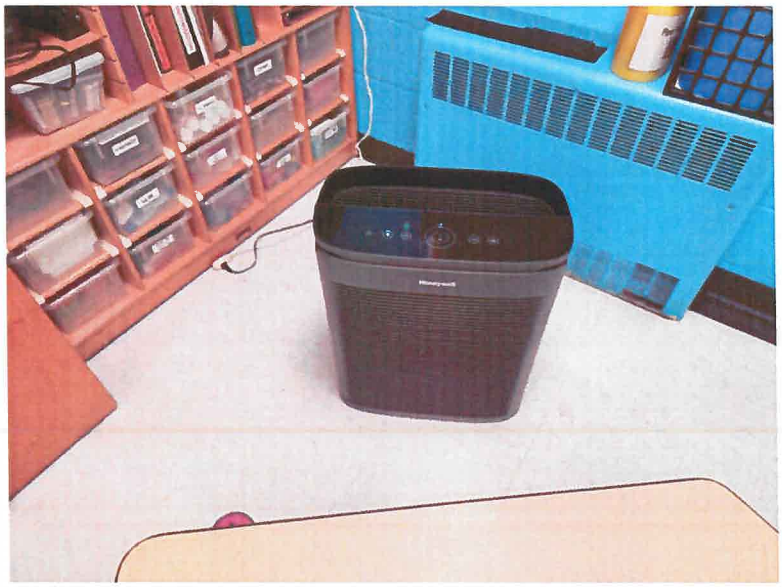

Photo #6:	View of Air Cleaner Operating in Classroom #5

Mid-Atlantic Environmental Consultants

PHOTOGRAPHIC RECORD

SALEM UNITED METHODIST CHURCH - CLASSROOM #5

Photo #7:	View of Air Cleaner Operating in Classroom #5

Appendix C – Accreditation

American Council for Accredited Certification

hereby certifies that

Joseph D. Pillart, Jr.

has met all the specific standards and qualifications of the re-certification process, including continued professional development, and is hereby re-certified as a

CIE

Council-certified Indoor Environmentalist

This certificate expires on May 31, 2024.

Charles F. Wiles, Executive Director

00859

Certificate Number